

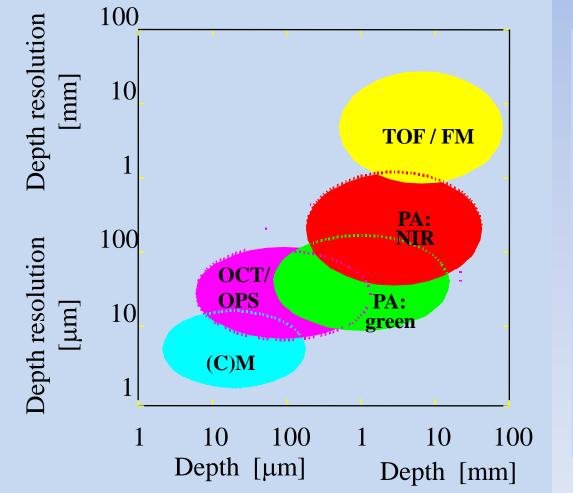
Photoacoustic Imaging of Blood Vessels in Tissue

F.F.M. de Mul

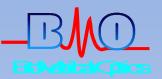
(University of Twente, Enschede, the Netherlands)

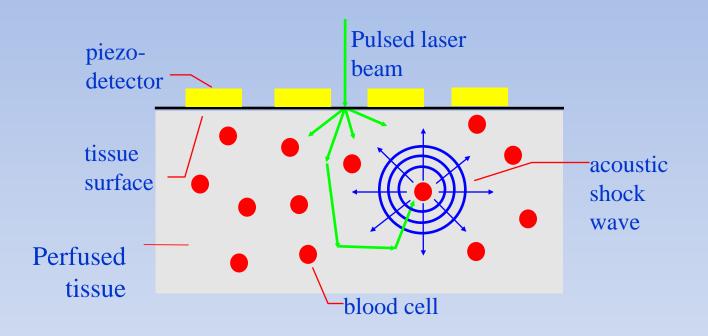


Imaging methods for hidden structures in turbid media (tissue)



C(M) : (confocal) microscopy **OCT:** optical coherence tomography **OPS:** orthogonal polarization spectral imaging PA: photoacoustics TOF: time-of-flight tomography FM: frequencymodulated tomography





- light pulse is absorbed in blood cell
- adiabatic heating
- pressure pulse emerging ($\approx 1500 \text{ m/s}$)
- detection at tissue surface

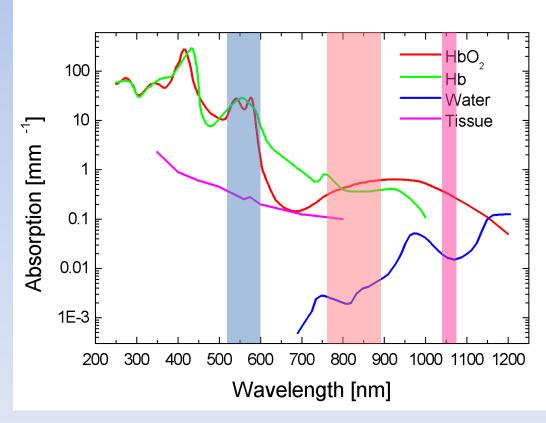
Depth:

- Green light: $\approx 0 9 \text{ mm}$
- Near-infrared: $\approx 0 30 \text{ mm}$

Depth resolution: $\approx 10 \ \mu m$

- BMD Optical properties of tissue and blood

Scattering and absorption



(Reduced) Scattering coefficient:

• $\lambda = 580$ nm:

Dermis: 3 mm⁻¹

- Blood: 1 ...
- λ = 850 nm:
 Dermis: 1 ...
 Blood: 0.5 ...

Available windows:

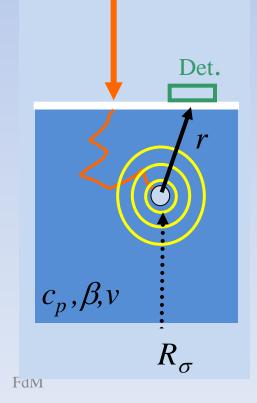


Photoacoustic Signal Generation

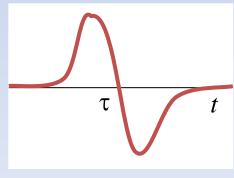
Bipolar PA-signal generated by a spherical Gaussian Source

$$\tau_a = \frac{R_\sigma}{\nu} \qquad \qquad \tau_e = \sqrt{\tau_a^2 + \tau_l^2}$$

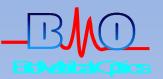
$$P(r,t) = -P_{\max}(r)\sqrt{e} \frac{t-\tau}{\tau_{e}} \exp\left\{-\frac{1}{2} \frac{(t-\tau)^{2}}{\tau_{e}^{2}}\right\}$$

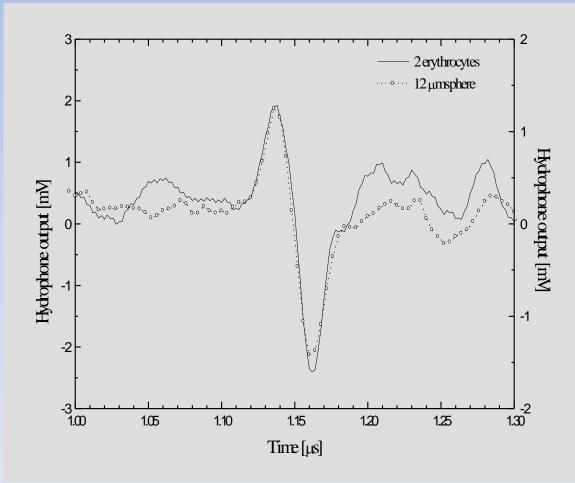


 E_a , τ_l



 $P_{\max}(r) = \frac{\beta E_{a}}{2\sqrt{e}(2\pi)^{3/2} c_{n} \tau_{e}^{2} r}$

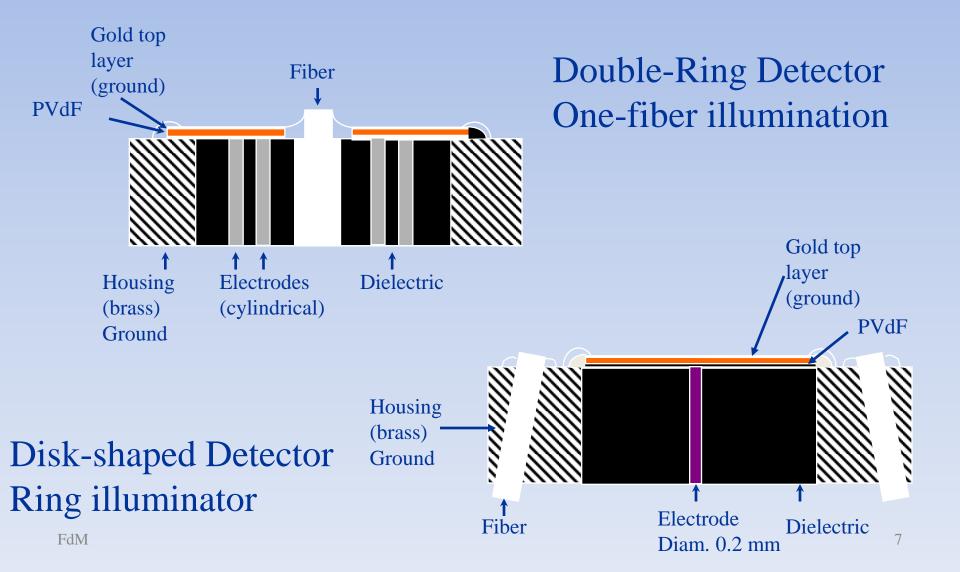


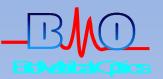


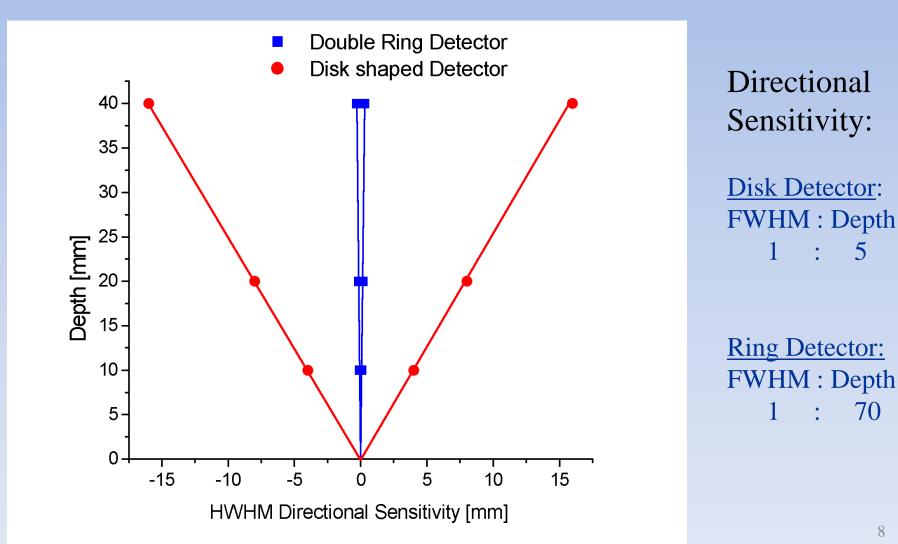
Two erythrocytes

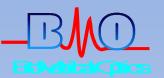
Diameter: $\approx 10 \ \mu m$ (compared with a 12 μm blue polystyrene sphere) detection distance: \approx 1.7 mm (= 1.15 $\mu s x$ 1500 m/s) medium: water/PBS

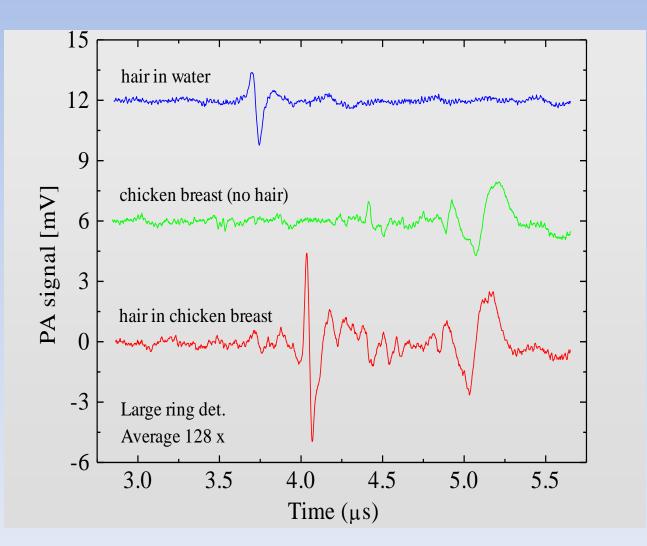










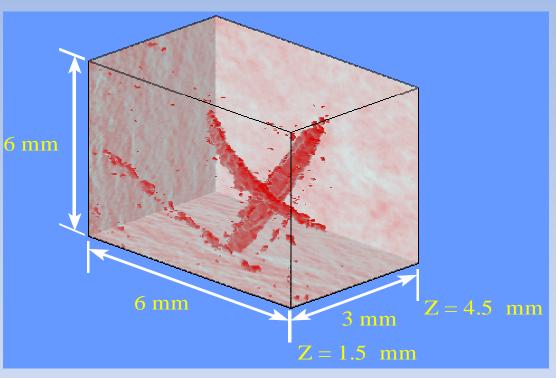


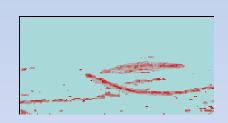
A human hair in chicken breast tissue.

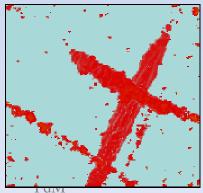
Depth: ≈ 6 mm (≈ 4 µs x 1500 m/s)

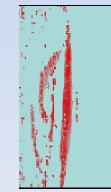


Vessels in chicken breast tissue



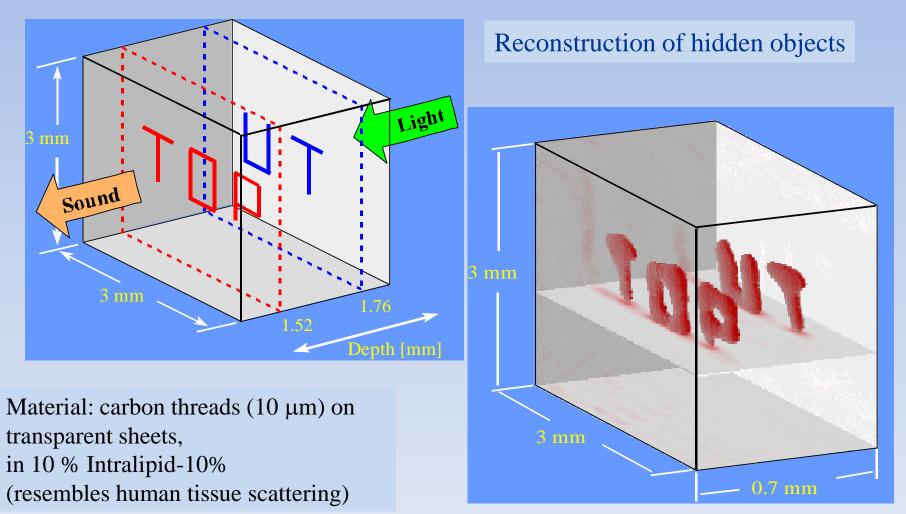






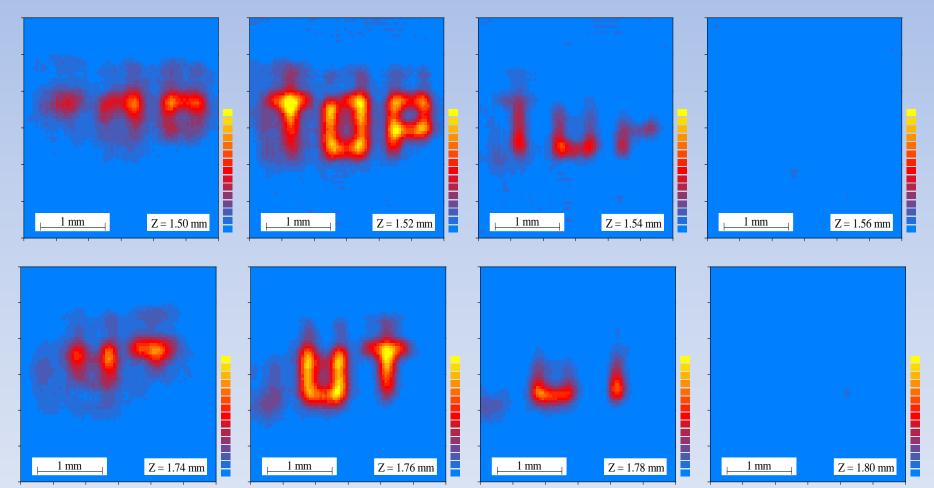
Sample : 5 mm thick chicken breast tissue in water Image : 663 mm, inside sample, 35 % isosurface threshold Vessels : 3 Nylon capillaries, 0.28 0.40 mm diameter Absorber : Evans Blue, flowing, a = 300 cm-1 Detection : at Z = 0 mm, 51 51 points, 0.15 mm spacing



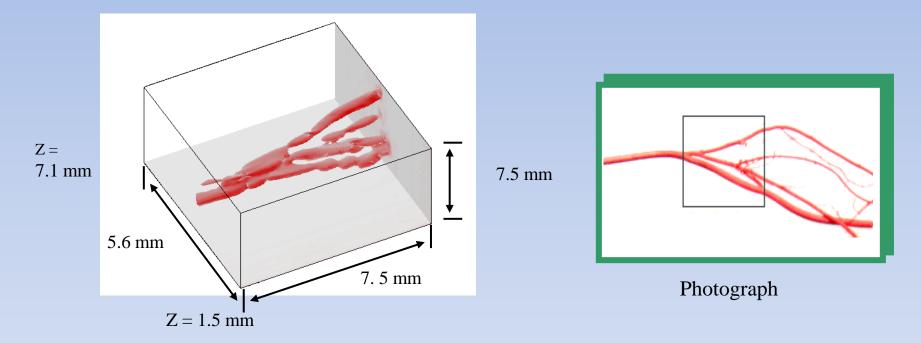




Depth resolution: $\approx 10 \,\mu$ m

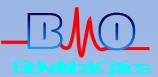






Vascular tree from a branching epigastric artery of a rat. Ex-vivo; medium: intralipid 1 % (\approx tissue). Depth (Z-coord.) \approx 5 mm : indicated in figure. Laser power 532 nm, 2mJ/pulse through fiber Ø 600 µm. Depth resolution / lateral resolution: 10 / 100 µm respectively.

FdM



PA-imaging of blood vessels in tissue

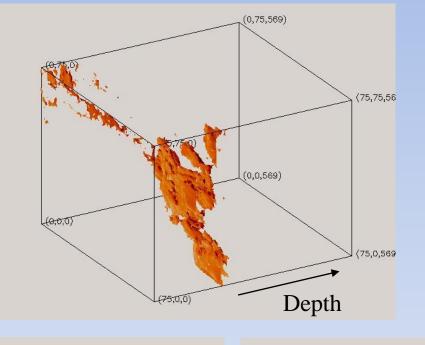
Live Albino Wistar Rat: Epigastric artery branching

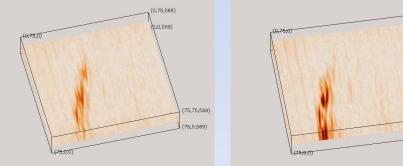
0.75.569

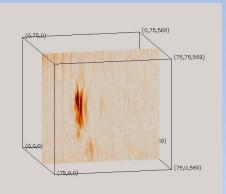
,0,569)

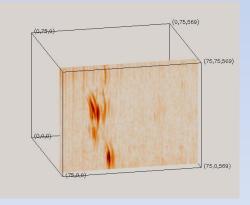
75.75.569

75.0.569

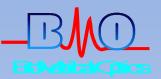








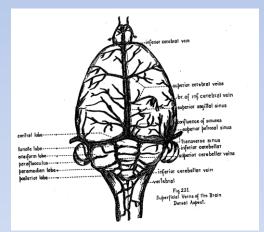
 $7.5 \text{ x } 7.5 \text{ x } 7.5 \text{ mm}^3$

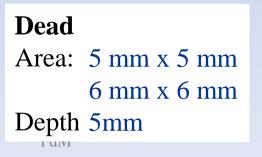


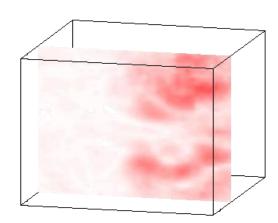
PA-imaging of blood vessels in tissue

Albino Wistar Rat: Brain Perfusion

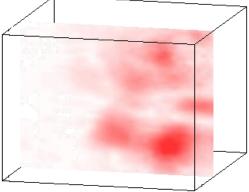
Alive Area: 4 mm x 4 mm Depth 5mm



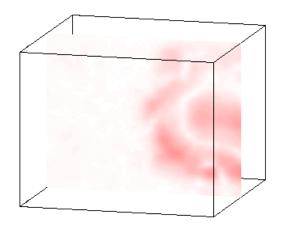


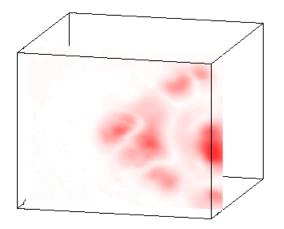


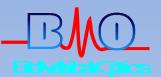
Slices of 100 µm



Blood in the cartilage scalp, in the upper part of the occipital bone

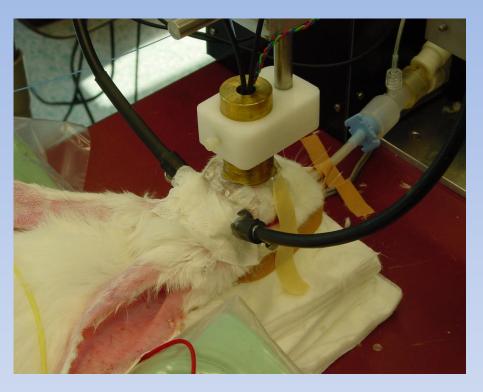






PA-image of Vessels in Rabbit Ear

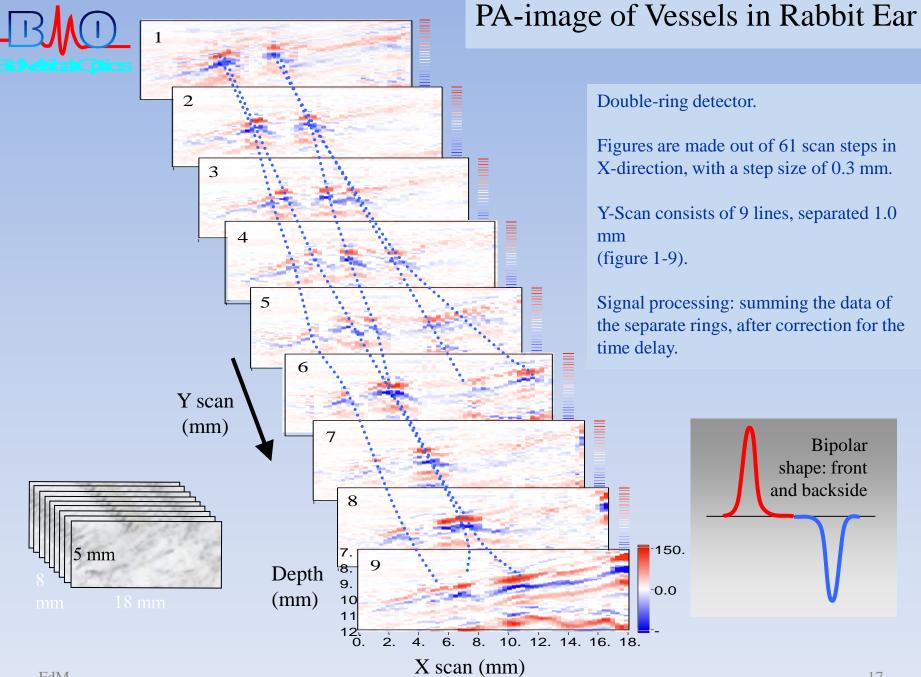


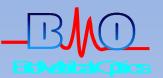


Double-ring detector.

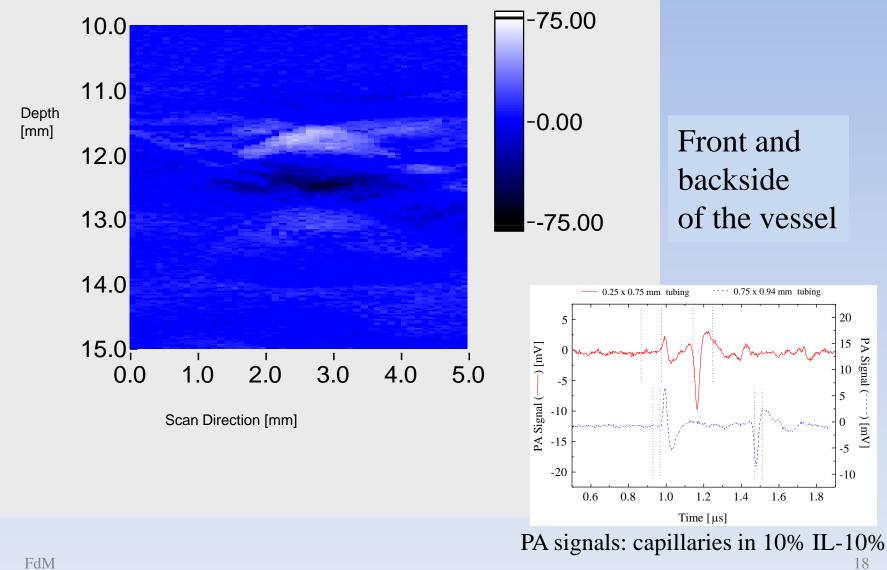
Figures are made out of 61 scan steps in X-direction, with a step size of 0.3 mm. Y-Scan consists of 9 lines, separated 1.0 mm

Signal processing: summing the data of the separate rings, after correction for the time delay.

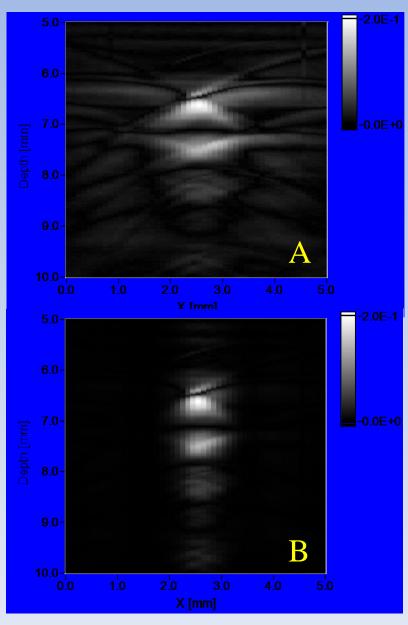




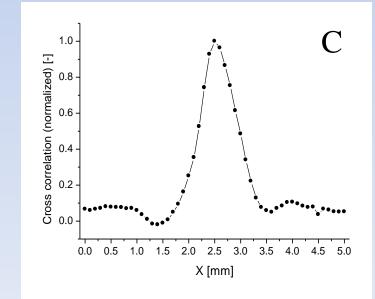
PA-imaging of blood vessels in rabbit ear



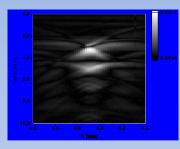
PA-imaging of blood vessels: image correction



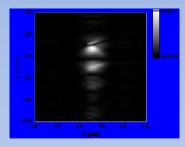
- 1mm diameter rubber tube, filled with human blood, in 7.5% Intralipid-10% dilution.
- Image A : 2D image, from measured time signals (1D-depth images), plotted as lines in the 2D image next to each other.
- Image B : 2D image, after correction by multiplication with the zero-time cross-correlation function, (C): <u>STEP 1</u>



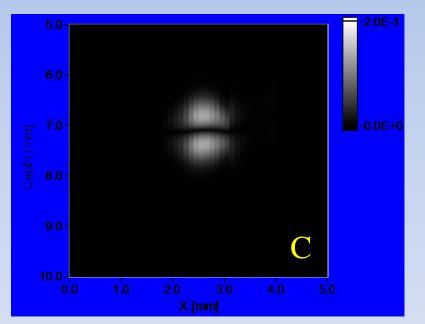
() PA-imaging of blood vessels: image correction



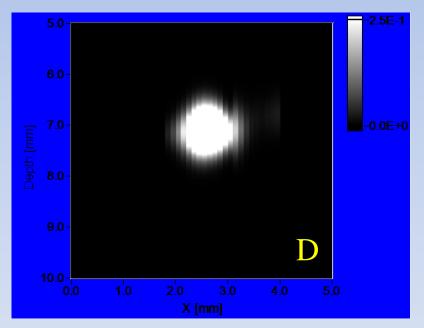
A: originally measured time signals



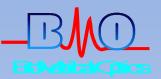
B: after multiplication with autocorrelation (STEP 1)



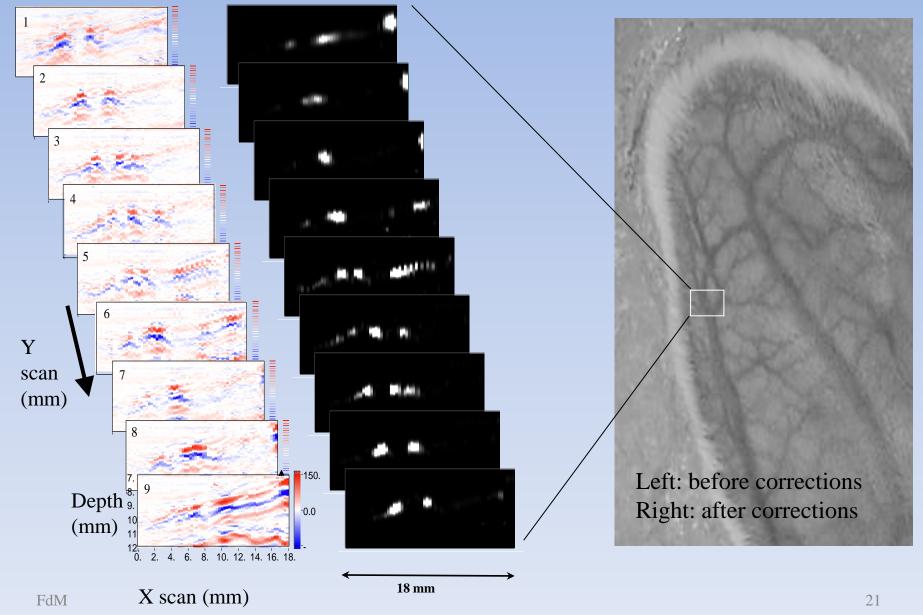
C: after fit to bipolar signal (STEP 2) Upper/lower image: pos./neg. peak signal (here absolute value plotted)

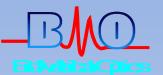


D: after running integration over depth (STEP 3)

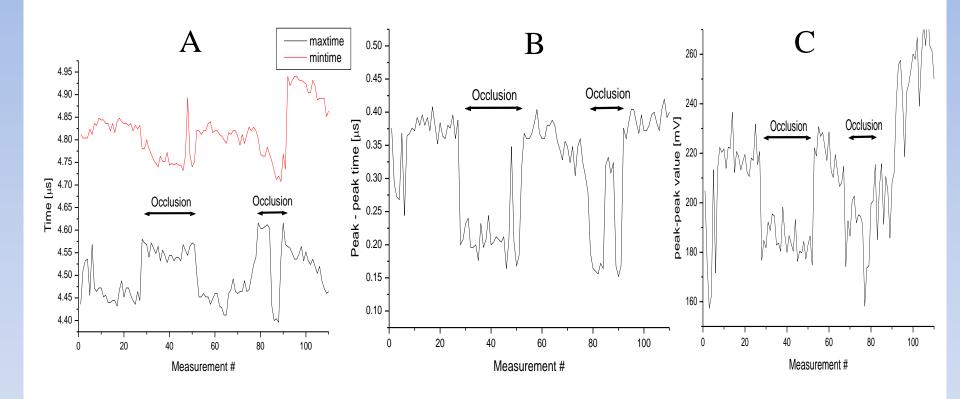


PA-imaging of blood vessels in rabbit ear

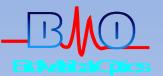




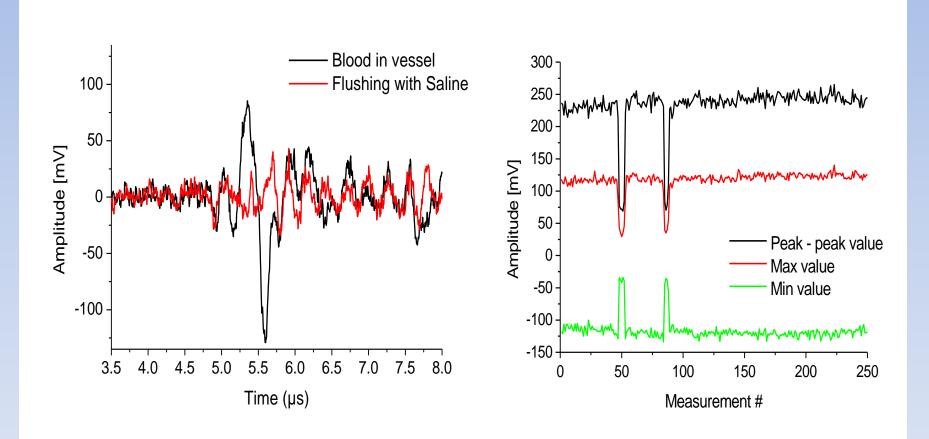
PA-imaging of blood vessels in rabbit ear: occlusion



During occlusion: A: Time between max / min peak reduced ($0.1 \ \mu s \approx 0.2 \ mm)$ B: Peak-peak time delay reduced => vessel thinner C: Peak-peak amplitude reduced => amount of blood lower



PA-imaging of blood vessels in rabbit ear: flushing with saline

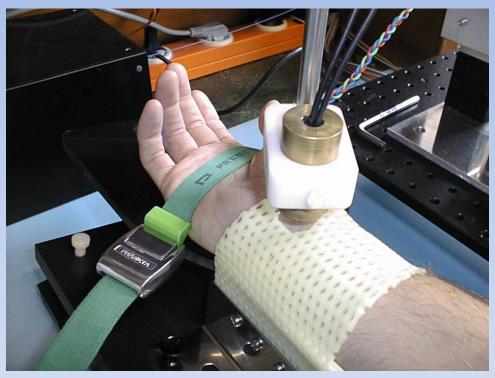




In vivo Scan: Human Blood Vessels



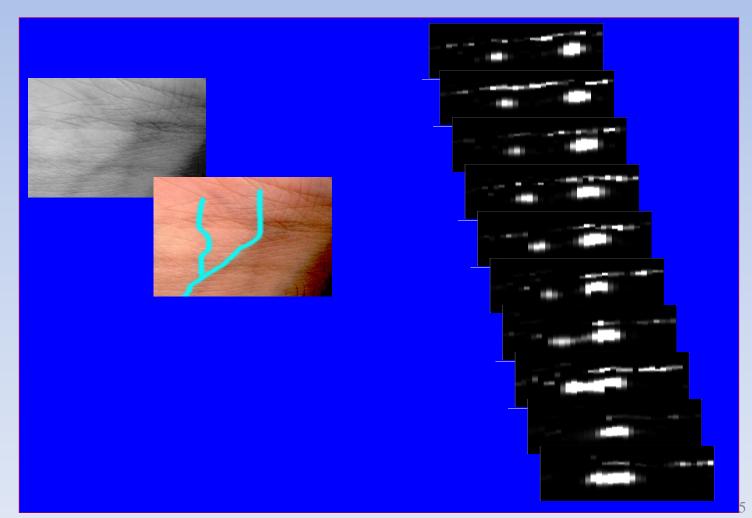




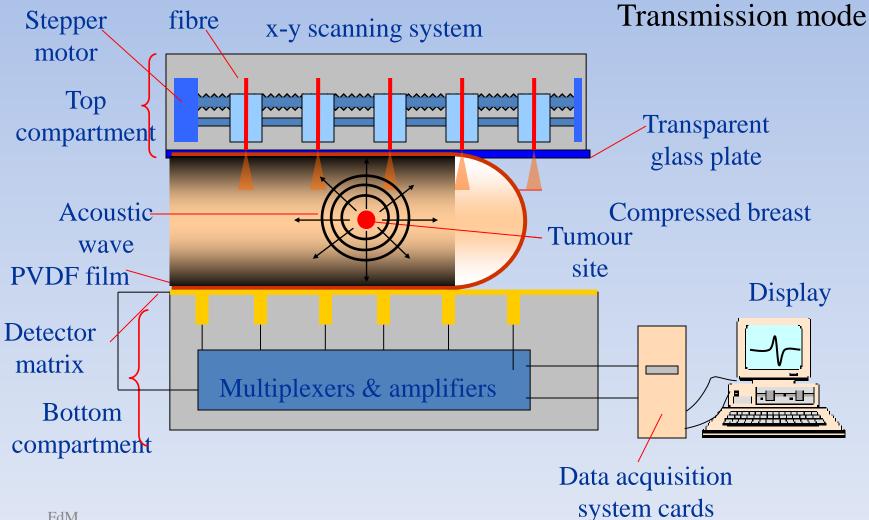


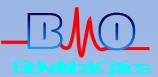
PA-imaging of blood vessels in human arm

Depth \approx 1-3 mm

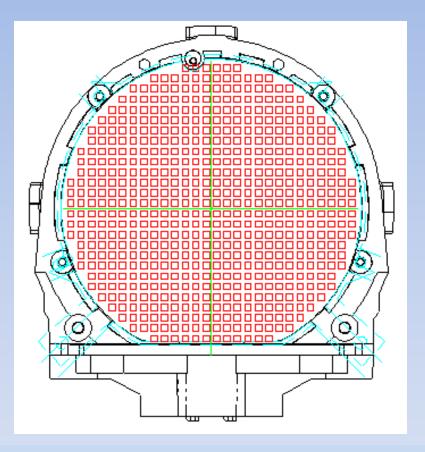


Photoacoustic Mammography

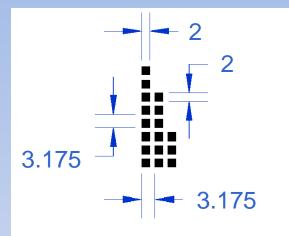




The detector matrix

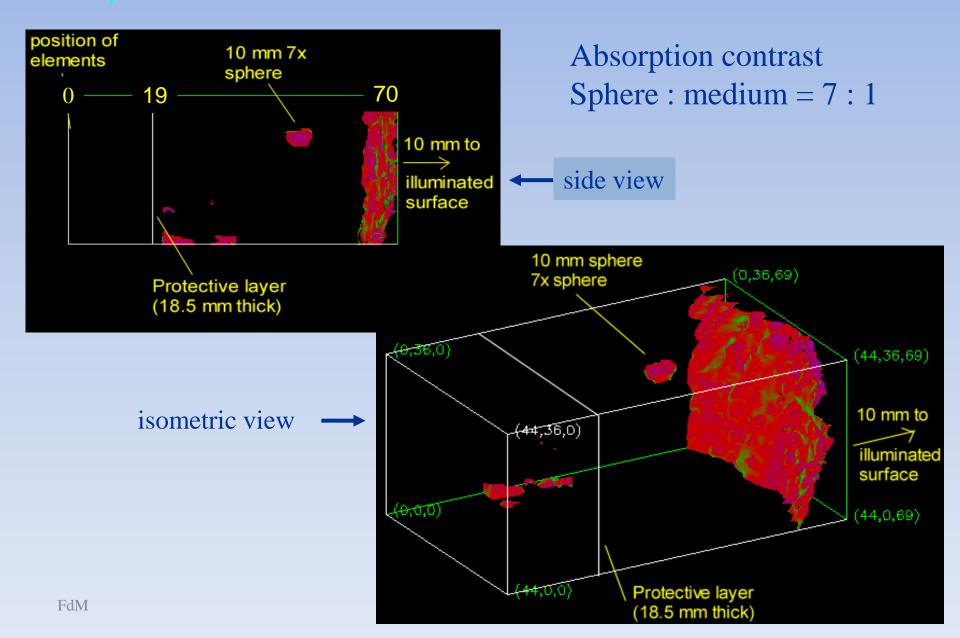


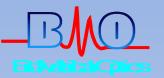
- 590 element microphone matrix
- active area: 90 x 85 mm



- arranged in a cartesian grid
- elements size: 2x2 mm
- separation: 3.175 mm
- 1 element accessed at a time



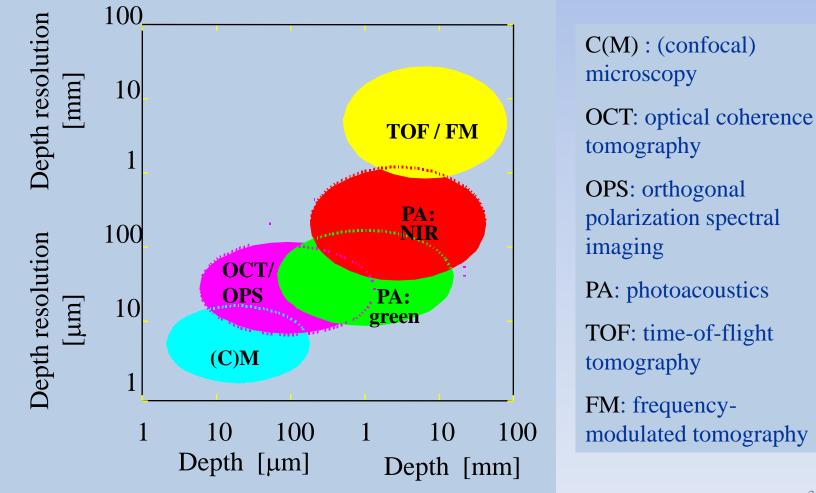


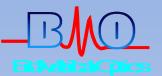


	Green 550 nm		NIR 800-1064 nm	
	dermis	blood	dermis	blood
 Absorption coefficient [1/mm] Scattering coefficient 	0.03	32	0.01	1.2
(reduced) [1/mm]	3	1	1	0.5
Absorption Contrast	1000		100	
 Penetration into tissue [mm] 	≈ 10		≈ 30	
 Applications 	Cutaneous perfusion Wound healing Diabetes Vascular malformations Skin tumours		Cerebral perfusion Muscular perfusion Mammography Angiogenese	



Imaging methods for hidden structures in turbid media (tissue)





Photoacoustic Imaging Of Blood Vessels in Tissue

The end